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DIPSLIDE CULTURE TEST

DIRECTIONS FOR USING DIPSLIDE

1. Unscrew the cap and withdraw the cap/slide from the vial. Be careful not to touch the agar coated surface of the slide.
2. Dip the slide into the fluid to be tested, covering the agar surface for a minimum of three seconds before removing.
3. Allow excess fluid to drain from the slide.
4. Screw the cap back on lightly and then back it off one half turn. Incubate the vial in an upright position at 25-36°C (77-96.8°F) depending on organism of interest and in-house protocol. The slide should be examined in the plastic vial for growth 24 hours after commencement of incubation and after a total of 48 hours for bacterial growth. If the dipslide is being used for fungal evaluation, the slide should be examined after 48 hours incubation and daily thereafter for a total of five days.

INTERPRETATION OF RESULTS

A. Bacteria (Nutrient agar – transparent media)

Bacteria grow either as red or colourless colonies. To determine the number of colony forming units compare the colony density on the transparent side with the density on the bacterial chart (Figure 1). Colourless colonies should be included in the comparison. It is the number of colonies and not the size of colonies that is important. Microbial counts in excess of 10^7 may appear as a uniform pink or red layer. A dilution is required to obtain an accurate count on such samples.

SAMPLE DILUTION INSTRUCTIONS

A sample may be diluted by adding 1 ml sample to 99 ml of boiled cooled tap water in a clean container. Cap the container and shake vigorously. Then, dip the slide into the diluted sample and proceed as above. Take the dilution factor into account when estimating microbial counts. For example, a 10^5 count from a 1/100 dilution of fluid would indicate 10^7 CFU/ml original sample.

B. Mold and Yeasts (Malt extract (brown) or Rose Bengal (pink) media)

The brown or pink side will detect the presence of yeasts and/or mould. Yeasts grow as smooth round colonies and mold as fuzzy colonies. Growth on this side may consist of pure yeast or mold or a mixture of both. Growth and type should be recorded when first seen, but incubation should be continued for 4 to 5 days to obtain good evaluation of possible fungal contamination.
To estimate yeast levels, compare yeast growth on fungal side to yeast colony density chart (see Figure 1). Any fungal growth in excess of one or two colonies per slide requires corrective action.

STORAGE

Dipslides should be stored at room temperature (20-25°C) (68-77°F) and protected from light. DO NOT REFRIGERATE OR FREEZE.

DISPOSAL OF EXPOSED SLIDES

Autoclave or incinerate the capped vials (caps loosened to prevent explosion) or open them under a fume hood and fill them with bleach. The bleach filled vials should stand overnight and then be disposed of in a prudent manner.

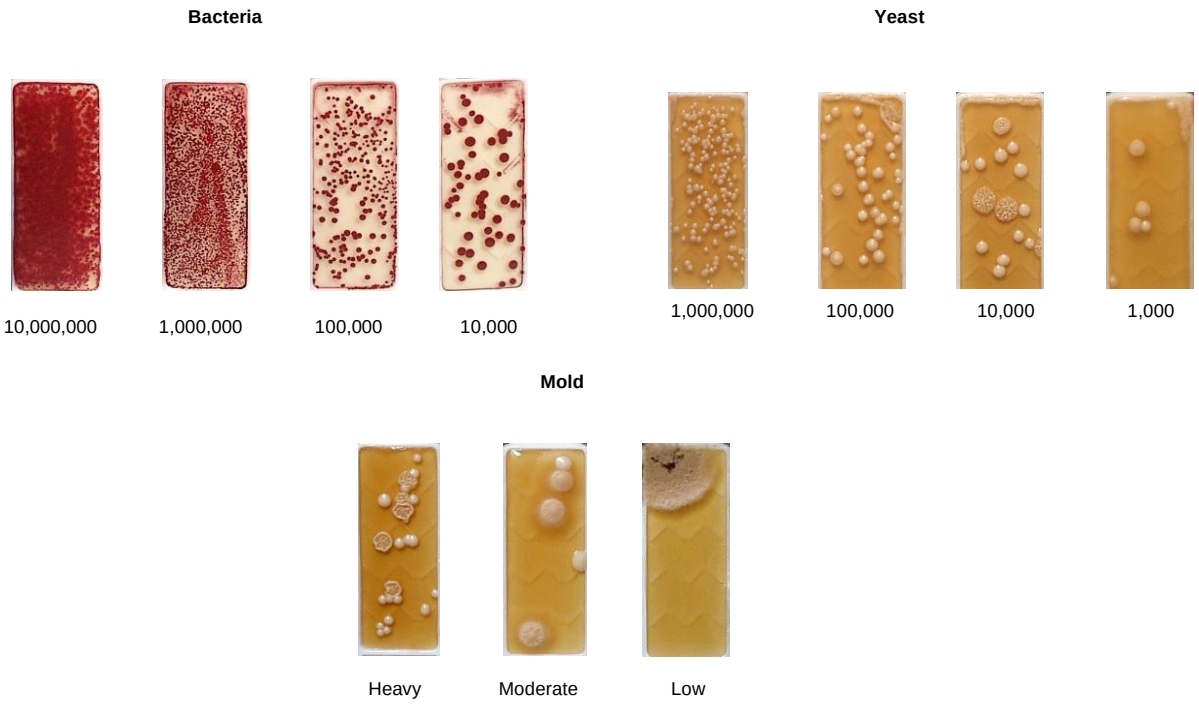

Figure 1. Colony density chart. Values shown are colony forming units per milliliter.
(Yeast and Mold dipslide with Rose Bengal are pink)